IN THE SPECIFICATION:

Please replace paragraph **[0110]** with the following amended paragraph:

[0110] Plasmids: The full-size mouse SIP1 cDNA sequence was cloned into the Myc-tag containing pCS3 eukaryotic expressing vector derived from pCS2 (69). The resulting plasmid was designated "pCS3-SIP1FS". Remacle et al. (68) described mutagenesis of the zinc finger clusters of the SIP1. For the construction of the inducible vector pUHD10.3SIP1, a *ClaI*/*Xba*I fragment from pCS3SIP1FS was cloned into the *EcoRI*/*Xba*I-cut pUHD10.3 vector (79). The *ClaI* site of SIP1 fragment and the EcoRI site of the vector were blunted using Pfu polymerase (Stratagene; La Jolla, CA). The E-cadherin promoter sequence (-341/+41) was obtained by PCR on genomic DNA from the human MCF7/AZ cell line. PCR-primers used are: 5'-ACAAAAGAACTCAGCCAAGTG-3' (SEQ ID NO:42) and 5'-CCGCAAGCTCACAGGTGC-3' (SEQ ID NO:43). The GC-melt kit (Clontech; Palo Alto, CA) was used for efficient amplification. The PCR product was blunted, kinased and then cloned into the pGL3basic vector (Promega; Madison, WI), which was opened at the *Srf*I site. By using the *Kpn*I-*Hind*III sites in this luciferase reporter construct, the E-cadherin promoter was also transferred to the pGL3enhancer vector. Mutagenesis of the E-boxes in the human E-cadherin promoter was performed by the ~~QuickChange~~ QUIKCHANGE™ Site-Directed Mutagenesis Kit (Stratagene) using the following primers:

forward primer E-box1: 5'-gctgtggccggCAG**A**TGaaccctcag-3' (SEQ ID NO:44);

reverse primer E-box1 : 5'-ctgagggttCA**T**CTGccggccacagc-3' (SEQ ID NO:45);

forward primer E-box3 : 5'-gctccgggctCA**T**CTGgctgcagc-3' (SEQ ID NO:46);

reverse primer E-box3 : 5'-gctgcagcCAG**A**TGagccccggagc-3' (SEQ ID NO:47).

Please replace paragraph **[0112]** with the following amended paragraph:

[0112] Promoter reporter assays: MCF7/AZ cells were transiently transfected by using FuGENE 6 (Roche; Basel, CH). NMe and MDA-MB231 were transfected with the LIPOFECTAMINE (Gibco BRL; Rockville, MD) procedure and the parental MCDK cell line

was transiently transfected with LIPOFECTAMINEPLUS™ (Gibco BRL; Rockville, MD). For transient transfection, about 200,000 cells were seeded per 10-cm^2 well. After incubation for 24 h, 600 ng of each plasmid type DNA was transfected. The medium was refreshed 24 h after transfection. Cells were lysed after 3 days in GALACTO-STAR™ kit lysis solution (Tropix, Bedford, MA). Normalization of transfection was done by measuring β-galactosidase, encoded by the cotransfected pUT651 plasmid (Eurogentec; Seraing, BE). Luciferase substrate is added to each sample. For β-galactosidase detection, a chemiluminescent substrate is supplied (Tropix, Bedford, MA). Luciferase and β-galactosidase activity was assayed in a ~~Topcount~~ TOPCOUNT™ microplate scintillation reader (Packard Instrument Co., Meriden, CT).

Please replace paragraph **[0113]** with the following amended paragraph:

[0113] Northern analysis: Total RNA was isolated with the RNeasy kit (Qiagen; Chatsworth, CA) following the manufacturer's protocol. Total RNA (25 μg) was glyoxylated, size-fractionated on a 1% agarose gel and transferred onto a ~~Hybond~~ HYBOND™ -N^+ membrane (Amersham Pharmacia Biotech, Rainhalm, UK). Hybridizations were performed as described before (81). The mouse SIP1 probe (459 bp) was generated by an *EcoR*-I digest of the mouse SIP1 cDNA. The human SIP1 probe (707 bp) was created by a *Bst* EII-*Not*I digest on the Kiaa 0569 clone (Kazusa DNA Research Institute). The mouse E-cadherin probe used was a *Sac*I fragment (500 bp) of the mouse E-cadherin cDNA. Two degenerated primers: 5' CTTCCAGCAGCCCTACGAYCARGCNCA 3' (SEQ ID NO:48) and 5' GGGTGTGGGACCGGATRTGCATYTTNAT 3' (SEQ ID NO:49) were used to amplify a fragment of the dog Snail cDNA from a total cDNA population of the MDCK cell line. Cloning and sequencing of the amplified band revealed a 432 bp cDNA fragment. To control the amount of loaded RNA, a GAPDH probe was used on the same blot. We performed the quantification of the radioactive bands on a ~~Phosphor Imager~~ PHOSPHOR IMAGER™ 425 (BioRad, Richmond, CA).

Please replace the Sequence Listing beginning on page 55 of the specification with the Substitute Sequence Listing Sheets, provided as Exhibit A hereto.